# 6-HYDROXY- AND 6-METHOXYFLAVONOIDS FROM NEUROLAENA LOBATA AND N. MACROCEPHALA

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**Key Word Index**—Neurolaena lobata; N. macrocephala; Asteraceae; Heliantheae; Galinsogineae; 6-hydroxykaempferol methyl ethers, glucosides and sulfate; quercetagetin methyl ethers, glucosides and sulfate; 6-hydroxyflavone methyl ethers and glucoside.

Abstract—Twelve flavonoids including one new sulfate were isolated from Neurolaena lobata, and six known flavonoids were obtained from N. macrocephala. The new compound isolated from N. lobata is 6-hydroxykaempferol 3-methyl ether 7-sulfate, and the known flavonoids are 6-hydroxykaempferol 3,7-dimethyl ether, 6-hydroxykaempferol, 3-methyl ether 7-glucoside, 6-hydroxykaempferol 7-glucoside, quercetagetin and its 7-glucoside, quercetagetin 3,6- and 3,7-dimethyl ethers, quercetagetin 3-methyl ether 7-glucoside and 7-sulfate, 6-hydroxyluteolin 3'-methyl ether and 6-hydroxyluteolin 7-glucoside. The known flavonoids identified from N. macrocephala are quercetagetin 3,6- and 3,7-dimethyl ethers, quercetagetin 6-methyl ether 7-glucoside, quercetagetin 3,6-dimethyl ether 7-glucoside, quercetagetin 7-glucoside and quercetagetin 3-methyl ether 7-sulfate.

#### INTRODUCTION

In a continuation of our biochemical systematic investigation of the genus Neurolaena (Asteraceae-Heliantheae) [1, 2], we report the isolation and characterization of twelve flavonoids from N. lobata (L.) R. Br., a widespread tropical weed in southern Mexico and South America, and six from N. macrocephala Sch.-Bip. ex Hemsl., a taxon endemic to Veracruz, Mexico. One of the flavonoids from N. lobata, 6-hydroxykaempferol 3-methyl ether 7-sulfate, is a new compound. We previously described fifteen flavonoids from N. oaxacana B. L. Turner [1], and seven from N. venturana B. L. Turner [2]. Two sesquiterpene lactones were previously reported from N. lobata [3], and the thymol derivatives of N. lobata, N. oaxacana and N. venturana have been described [4].

### RESULTS

Leaves of N. lobata and N. macrocephala were both extracted with aqueous methanol and the syrup obtained after concentrating the extract was partitioned between water and three organic solvents: n-hexane, dichloromethane and ethyl acetate. The hexane extract was discarded. The dichloromethane extract of N. lobata yielded 6-hydroxykaempferol 3,7-dimethyl ether (1) [5], quercetagetin 3,7-dimethyl ether (2)[5], quercetagetin 3,6-dimethyl ether (3)[6], 6-hydroxyluteolin 3'-methyl ether (4) [7], and 6-hydroxykaempferol 7-glucoside (5) [8]. The ethyl acetate extract yielded additional 6hydroxykaempferol 7-glucoside plus quercetagetin (6) [9], and 6-hydroxyluteolin 7-glucoside (7) [10]. The water extract yielded the remaining compounds: quercetagetin 3-methyl ether 7-sulfate (8) [1], 6-hydroxykaempferol 3-methyl ether 7-sulfate (9), 6-hydroxykaempferol 3-methyl ether 7-glucoside (10)[1], quercetagetin 3methyl ether 7-glucoside (11)[1] and quercetagetin 7-glucoside (12)[11].

The identities of all the known flavonoids were determined by direct comparison (TLC, UV, NMR) with authentic samples previously obtained from *N. oaxacana* [1]. The structure assignment of the new compound is discussed separately, and all previously unreported color, TLC, UV, NMR and MS data for all the flavonoids are recorded in Tables 1 and 2 or in the Experimental.

6-Hydroxykaempferol 3-methyl ether 7-sulfate (9)

The presence of a 6-hydroxyl group in 9 was first suspected from the purple-brown color of the spot on a paper chromatogram under UV light which was unchanged when exposed to ammonia vapor or sprayed with NA. The UV band I shift of +24 nm in AlCl<sub>3</sub>/HCl relative to band I in MeOH indicated a flavonoid with C-6 oxygenation. The absence of a band III, band I at 390 nm in NaOMe vs 400 nm in NaOAc, and no shift of band II in NaOAc suggested an -OR substituent at C-7 [12]. The lack of a shift of band I with NaOAc/H<sub>3</sub>BO<sub>3</sub> established that there was no ortho-dihydroxyl group in the B-ring. The electrophoretic mobility to the anode (4.5 cm) of this compound under standard conditions [13] at pH 1.9 indicated that it was a monosulfated compound. This was confirmed when sulfatase hydrolysis yielded 6hydroxykaempferol 3-methyl ether (TLC comparison with an authentic sample). The UV spectral findings require that the sulfate moiety is at C-7. Thus, the new compound is 6-hydroxykaempferol 3-methyl ether 7sulfate.

The flavonoids isolated from the dichloromethane extract of *N. macrocephala* were quercetagetin 3,6-dimethyl ether (3)[6] and quercetagetin 3,7-dimethyl ether (2)[5]. The ethyl acetate extract yielded quercetagetin 6-methyl ether 7-glucoside (13)[14], quercetagetin

Table 1. Chromatographic data ( $R_s \times 100$  and colors) for flavonoids of N. lobata\*

				Cell	ulose						
	Pa	iper	НС	)Ac	ТВА	n-BAW	Poly	amide		Colors†	
Compound	ТВА	НОАс	15%	40 %	-		BMM	ВРММ	UV	UV/NH	UV/NA
6-Hydroxykaempferol											
7-glucoside (5)	25	9	5	28	19	24	29	0	PBr	PBr	PBr
Quercetagetin (6)	20	0.2	2	10	19	37	19	2	P	PBr	Or
6-Hydroxykaempferol 3-methyl ether 7-sulfate (9)	39	68	56	79	45	48		_	P	PBr	PBr
Quercetagetin 3,6-dimethyl ether	3,	-				.0			-		
7-glucoside (14)	44	36	34	57	36	40	0	0	P	YBr	Or

\* See ref. [1] for chromatographic data of other flavonoids from N. lobata and solvent key.

3,6-dimethyl ether 7-glucoside (14) [8], and quercetagetin 7-glucoside (12) [11]. Quercetagetin 3-methyl ether 7sulfate(8)[1] was obtained from the water fraction. The identities of all these flavonoids were determined by direct comparison (TLC, UV) with authentic samples previously obtained from N. oaxacana [1]. The UV and chromatographic data for quercetagetin 3,6-dimethyl ether 7-glucoside are reported in Tables 1 and 2.

## EXPERIMENTAL

Plant material. Leaves and vouchers of N. lobata were collected from along the roadside several km N. of Atzalan at La Calavera. Vercruz, Mexico, on 9 March 1979 (voucher specimen K. M. Kerr 124 is deposited in the Lundell Herbarium, The University of Texas at Austin). Leaves and vouchers of N. macrocephala were collected 300 m from the entrance to Los Tuxtlas Biological

 $R^1 = Me, R^2 = H$ 

 $R^1 = Me$ ,  $R^2 = OH$ 

 $R^1 = SO_3^2$ ,  $R^2 = H$ 

10  $R^1 = glc, R^2 = H$ 

 $R^1 = H, R^2 = Me$ 

7  $R^1 = glc, R^2 = H$ 

5  $R^1 = glc, R^2 = H$ 

6  $R^1 = H, R^2 = OH$ 

12  $R^1 = glc, R^2 = OH$ 

13

<sup>†</sup> P = purple, PBr = purplish-brown, Or = orange, YBr = yellowish brown; NA = Naturstoff reagenz A in MeOH.

Table 2. UV data  $(\lambda_{max}, nm)$  for flavonoids of N. lobata\*

Compound	МеОН	NaOMe	AICI <sub>3</sub>	AlCl <sub>3</sub> /HCl	NaOAc	NaOAc/H <sub>3</sub> BO <sub>3</sub>
5	374 (sh), 346 (1),†	400 (1), 290 (0.5),	436 (sh), 388 (1),	428 (1), 382 (1.2),	384 (sh), 348 (1),	380 (sh), 386 (1),
	276 (0.8),	254 (0.7)	287 (0.9)	267 (1.5)	272 (0.9),	278 (1.5)
	256 (0.7), 234 (0.7)				254 (0.8)	
9	360 (1), 272 (sh),	390 dec. (1),	444 (1), 280 (0.7)	436 (sh), 396 (1),	388 (1),	376 (1), 292 (sh),
	255 (0.7)	304 (3), 244 (4)		270 (1)	260 (0.7)	266 (0.7)
6	366 (1), 294 (sh),	390 (1), 300 (sh),	400 (sh), 364 (1),	398 (sh), 358 (1),	400 (sh),	
	270 (0.8)‡	270 (0.6)	304 (sh), 278 (1)	302 (sh), 280 (1.5)	344 (1), 268 (0.8)	366 (1), 270 (0.5)
14	350 (1), 292 (sh),	400 (1), 276 (sh),	434 (1), 364 (sh),	400 (sh), 370 (1),	370 (1), 290 (8),	370 (1), 290 (0.8), 264 (1.4)
	260 (1.5)	252 (1.3)	326 (sh), 304 (sh),	298 (sh),	260 (1.4)	
			280 (2)	268 (1)		

<sup>\*</sup> See ref. [1] for the UV data of other flavonoids from N. lobata.  $\dagger$  Relative absorptivities are given for each  $\lambda_{\max}$  relative to the most intense wavelength peak as (1),  $\dagger$  MeOH/HCI: 338 (1), 280 (0.8).

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Station on 13 March 1979 (voucher specimen K. M. Kerr 136 is deposited in the Lundell Herbarium, The University of Texas at Austin).

Extraction, purification and identification of flavonoids from N. lobata and N. macrocephala. General chromatographic and electrophoretic techniques have been previously described [1]. Ground leaves of N. lobata (200 g) were extracted  $3 \times$  with aq. MeOH. The combined extracts were concd in vacuo to 100 ml, and the aq. concentrate successively extracted with CH2Cl2 and EtOAc. A. CH<sub>2</sub>Cl<sub>2</sub> extract. The syrup from this extract (5 g) was passed over Sephadex LH-20, and the flavonoid mixture (0.7g) obtained from this column was chromatographed over a Polyclar column (4 × 17 cm; 50 g). Elution was initiated with Egger's solvent (CH<sub>2</sub>Cl<sub>2</sub>-MeOH-MeCOEt-Me<sub>2</sub>CO, 20:10:5:1) and the polarity of the solvent was gradually increased by the elimination of CH<sub>2</sub>Cl<sub>2</sub> (0:10:5:1). The compounds eluted in the following order: 6-hydroxykaempferol 3,7-dimethyl ether (1), 1.5 mg; quercetagetin 3,7-dimethyl ether (2), 3 mg; quercetagetin 3,6-dimethyl ether (3), 1 mg; 6-hydroxyluteolin 3'-methyl ether (4), 1 mg; and 6-hydroxykaempferol 7-glucoside (5), 14 mg. B. EtOAc extract. The syrup from this extract (9.2g) was passed over a Sephadex LH-20 column and the flavonoids were obtained as a mixture (3g) which was chromatographed over Polyclar  $(4 \times 32 \,\mathrm{cm}; 90 \,\mathrm{g})$ . The column was first eluted with H<sub>2</sub>O-MeOH-MeCOEt-Me<sub>2</sub>CO (13:3:3:1), and the polarity was decreased by the gradual elimination of H2O. The compounds obtained were quercetagetin (6), 5 mg, and 6hydroxyluteolin 7-glucoside (7), 8 mg. C. H<sub>2</sub>O extract. The aq. extract concentrate (11.7 g) was chromatographed over Polyclar  $(5 \times 35 \,\mathrm{cm}; 150 \,\mathrm{g})$  in the same manner as for the EtOAc extract. Compounds eluted in the following order: quercetagetin 3methyl ether 7-sulfate (8), 1 mg; 6-hydroxykaempferol 3-methyl ether 7-sulfate (9), 4 mg; 6-hydroxykaempferol 3-methyl ether 7glucoside (10), 14 mg; quercetagetin 3-methyl ether 7-glucoside (11), 11 mg; and quercetagetin 7-glucoside (12), 30 mg.

NMR and MS data for N. lobata flavonoids. 6-Hydroxy-kaempferol 7-glucoside (5) and quercetagetin (6); MS of 5 derivatized m/z (rel. int.), M<sup>+</sup> 302 (100), M – H 301 (40), M – H<sub>2</sub>O 286 (50), M – CHO 274 (60), M – COMe 259 (20), A<sub>1</sub> 168 (50), A<sub>1</sub> – 16 152 (60), B<sub>2</sub> 121 (100). <sup>1</sup>H NMR (90 MHz, CCl<sub>4</sub>, TMS):  $\delta$  3.3 – 3.8 (5 H, m, 6 glucosyl protons), 4.95 (1-H, d, J = 8 Hz, for H"-1), 6.5 (1 H, s, H-8), 6.8 (2 H, d, J = 9 Hz, H-3' and H-5'), 7.93 (2 H, d, J = 9 Hz, H-2' and H-6'). Compound 6: <sup>1</sup>H NMR (90 MHz, CCl<sub>4</sub>, TMS):  $\delta$  6.5 (1 H, s, H-8), 6.85 (1 H, s, s = 9 Hz, H-5'), 7.65 (1 H, s, s = 9 and 2 Hz, H-6'), 7.6 (1 H, s, s = 2 Hz, H-2').

The extraction techniques used for the isolation of flavonoids from 50 g of dry leaf material of *N. macrocephala* were similar to those already described for *N. lobata*. A. CH<sub>2</sub>Cl<sub>2</sub> extract. The syrup from the CH<sub>2</sub>Cl<sub>2</sub> extract was chromatographed by 1 D PC

in TBA (t-butanol-HOAc-H<sub>2</sub>O, 3:1:1) on Whatman 3MM paper. The flavonoids were eluted with MeOH and purified over Sephadex LH-20. The following compounds were obtained: quercetagetin 3,6-dimethyl ether (3), 1 mg, and quercetagetin 3,7-dimethyl ether (2),2 mg. B. EtOAc extract. After the same purification procedure of the conc. EtOAc extract the following compounds were isolated: quercetagetin 6-methyl ether 7-glucoside (13), 2 mg; quercetagetin 3,6-dimethyl ether 7-glucoside (14), 4 mg, and quercetagetin 7-glucoside (12), 5 mg. C. H<sub>2</sub>O extract. On purification as above the conc H<sub>2</sub>O extract gave quercetagetin 6-methyl ether 7-glucoside (13), quercetagetin 7-glucoside (12) and a trace of quercetagetin 3-methyl ether 7-sulfate (8).

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#### REFERENCES

- Ulubelen, A., Kerr, K. M. and Mabry, T. J. (1980) *Phytochemistry* 19, 1761.
- 2. Ulubelen, A., Kerr, K. and Mabry, T. J. (1980) *Planta Med.* (in press).
- Manchand, P. S. and Blount, J. F. (1978) J. Org. Chem. 43, 4352.
- 4. Bohlmann, F., Natu, A. A. and Kerr, K. (1979) Phytochemistry 18, 489.
- Shen, M. C., Rodriguez, E., Kerr, K. and Mabry, T. J. (1976) Phytochemistry 15, 1045.
- Dillon, M. O., Mabry, T. J., Besson, E., Bouillant, M. L. and Chopin, J. (1976) Phytochemistry 15, 1085.
- 7. Taylor, A. O. and Wong, E. (1965) Tetrahedron Letters 3675.
- Bacon, J. D., Urbatsch, L. E., Bragg, L. H., Mabry, T. J., Neuman, P. and Jackson, D. W. (1978) *Phytochemistry* 17, 1939.
- 9. Baker, W., Nodzu, R. and Robinson, R. (1929) J. Chem. Soc.
- Barua, A. K., Chadhabarti, P. and Sahyal, P. K. (1969) J. Indian Chem. Soc. 46, 271.
- Geissman, T. A. and Steelink, C. (1957) J. Org. Chem. 22, 946.
- Bacon, J. D., Mabry, T. J. and Mears, J. A. (1976) Rev. Latinoam. Ouim. 7, 83.
- 13. Al-Khubazi, M. (1977) M. S. Thesis, The University of
- Sharma, R. C., Zaman, A. and Kidwai, A. R. (1964) Indian J. Chem. 3, 83.